

## Presence of *Batrachochytrium dendrobatidis* at the Headwaters of the Mississippi River, Itasca State Park, Minnesota, USA

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The disease caused by the pathogenic fungus *Batrachochytrium dendrobatidis* (*Bd*), chytridiomycosis, is one of several factors driving the global decline of amphibian populations (Blaustein and Kiesecker 2002; Lips et al. 2006; Muths et al. 2003). *Bd* has been found in amphibians at sites across North America including Minnesota, USA (Ouellet et al. 2005; Woodhams et al. 2008). The prevalence of *Bd* in wild anuran populations in Minnesota is unknown, and motivated the work described herein.

We investigated the occurrence of *Bd* at the University of Minnesota's Itasca Biological Station and Laboratories in Itasca State Park, site of the headwaters of the Mississippi River. Our research objectives were to: 1) verify if *Bd* is present in the park; 2) determine which anuran species are affected by the fungus; and 3) test if there are differences in infection rate among species.

**Methods.**—We collected frogs in Itasca State Park, Clearwater County, Minnesota in June and July 2008. We collected frogs by hand and with nets at night in breeding ponds and during the day near ponds and wetlands. We rinsed nets with 95% ethanol between outings and wore latex gloves in the field and the laboratory to prevent potential transfer of *Bd* among individuals. A subsample of frogs was toe-clipped and released in the field, but most were vouchered. Toe-clips were collected in individual plastic bags and we extracted genomic DNA immediately upon return to the laboratory. We kept frogs individually in plastic bags from time of capture until they were euthanized in the laboratory. Frogs were humanely euthanized with MS-222 (tricaine methanesulfonate) or topical application of benzocaine (Simmons 2002). MS-222 does not appear to inhibit growth or detection of *Bd* (Webb et al. 2005) and was the preferred method of euthanasia. We clipped one toe and a portion of adjacent webbing to obtain tissues. We stored tissues in 95% ethanol at 4°C until processing. All vouchers were deposited in the Bell Museum of Natural History, University of Minnesota (JFBM).

We extracted genomic DNA from tissues using the Qiagen extraction kit (Qiagen, Valencia, CA, USA) following the manufacturer's

instructions. We used polymerase chain reaction (PCR) to amplify a 300-bp fragment consisting of part of internal transcribed spacer 1 (ITS1), ribosomal rRNA 5.8S, and part of internal transcribed spacer 2 (ITS2) using *B. dendrobatidis* specific primers (Bd1a and Bd2a; Annis et al. 2004). PCR was performed in 12.5 µl reaction volumes under the following conditions: an initial denaturation of 94°C for 5 min; 30 cycles of denaturation (94°C for 45 sec), annealing (50°C for 45 sec) and extension (72°C for 1 min); followed by a final extension of 72°C for 5 min. All PCR reactions contained a negative control. PCR products were run on a 1% agarose gel, stained with ethidium bromide and visualized under ultraviolet light. Presence of a strong, defined band approximately 300 bp was considered a positive result.

We sequenced two positive samples to ensure that we had amplified *Bd* rather than non-target DNA. We purified PCR products using Exonuclease I and Shrimp Alkaline Phosphatase (Hanke and Wink 1994). Sequencing was performed using Big Dye (Perkin Elmer, Boston, MA, USA) terminator cycle sequencing on an ABI 3730xl at the Advanced Genetic Analysis Center, University of Minnesota. We used BLAST (Altschul et al. 1990) to verify that sequences were *Bd*.

We tested whether infection rates of *Bd* were significantly different among sampled species and among sampled families of frogs using a Chi-square contingency test. All statistical analyses

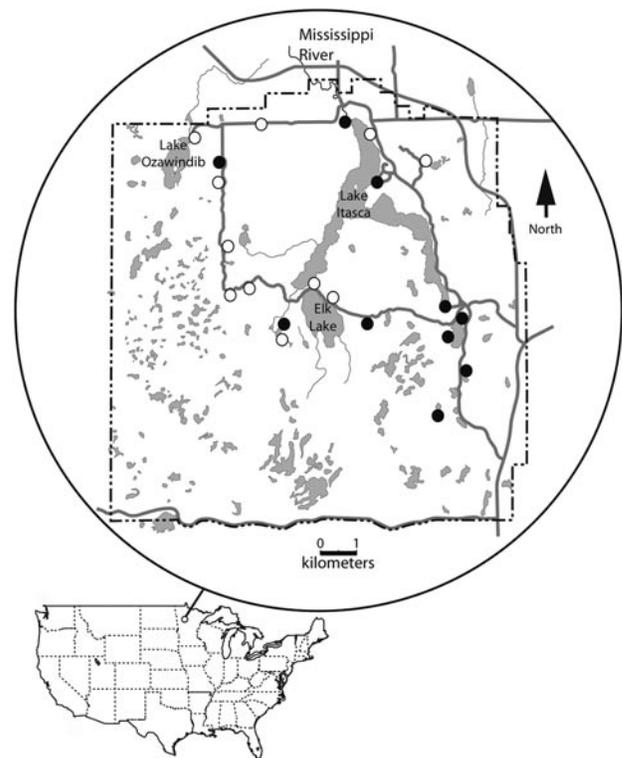


FIG. 1. Collecting locations (circles) of anurans examined for *Batrachochytrium dendrobatidis* (*Bd*) at Itasca State Park, Minnesota, USA. Localities with *Bd*-positive specimens indicated by filled circles, localities with no *Bd*-positive specimens indicated by open circles. Park boundaries shown by a dashed line. Light gray areas are water.

were conducted using JMP 7.0 (SAS 2007).

**Results.**—We collected tissues from 147 frogs and toads of three families from around Itasca State Park (Fig. 1), consisting of 133 vouchered specimens and 14 toe-clips of released frogs. Thirty-four of 147 (23%) individuals were *Bd*-positive. None of our negative controls produced bands on agarose gels. *Pseudacris maculata* had the highest percentage of positive individuals,

50% (Table 1), although it is difficult to interpret this given the small number of individuals captured. *Lithobates pipiens* had the second highest infection rate with 33.3% of samples being *Bd*-positive. *Hyla versicolor* and *Pseudacris crucifer* had no positive samples. We collected one dead *Lithobates sylvaticus* (JFBM 15957) at the Mississippi River headwaters on the North end of Lake Itasca that tested positive for *Bd*. No other dead animals were found, and no animals were observed with symptoms of illness.

Rates of *Bd* infection differed among species ( $\chi^2 = 16.505$ ,  $P < 0.0113$ ) and among families ( $\chi^2 = 9.673$ ,  $P < 0.0079$ ). Frogs in the family Ranidae had a higher infection rate than Bufonidae and Hylidae (Table 1).

We confirmed that DNA fragments amplified using PCR were *Bd* by sequencing two of our positive PCR products. Both sequenced samples, one from *Lithobates sylvaticus* (JFBM 15884, Genbank accession number FJ229469) and the other from *Lithobates pipiens* (JFBM 15918, Genbank accession number FJ229470) had an identical sequence. We compared our sequences to *Bd* sequences on Genbank using BLAST, confirming our samples as *Bd*. Our samples had 100% identity with the following *Bd* sequences from Genbank: EU779867, EU779864, EU779863, EU779862, EU779860, EU779859. Our samples had a 98% identity with *Bd* sequences AY997031 and EU779866.

**Discussion.**—Rates of *Bd* infection in Itasca State Park varied among species and families. This variance is similar to observations from other North American sites for the same species (Longcore et al. 2007; Ouellet et al. 2005). North American hylid frogs, for example, typically have had low *Bd* infection rates (Longcore et al. 2007; Ouellet et al. 2005; Pearl et al. 2007) and we found no evidence of *Bd* in the hylid species *Hyla versicolor* and *Pseudacris crucifer*. *Pseudacris maculata* and *Acris blanchardi* appear to be exceptions to this rule (Ouellet et al. 2005; Steiner and Lehtinen 2008) and we found one infected *P. maculata* in the Park. We found the highest rates of infection in the three ranid species examined, *L. sylvaticus*, *L. pipiens* and *L. septentrionalis*. Several hypotheses have been proposed to explain the species-specific variance in *Bd* infection rates. Because *Bd* can persist in aquatic environments (Johnson and Speare 2003), species that spend more time in the water are thought to be at greater risk of infection than species spending less time in the water (Hero et al. 2005; Lips et al. 2003). *Bd* is prevalent in species breeding in permanent wetlands and streams (Kriger and Hero 2007) and in species that overwinter in aquatic environments (Longcore et al. 2007). There is also, poten-

TABLE 1. Prevalence of *Batrachochytrium dendrobatidis* in seven frog species collected in Itasca State Park, Minnesota, USA, in 2008.

Species	Family	No. <i>Bd</i> - Negative Animals	No. <i>Bd</i> -Positive Animals	% Positive (95% C.I.)
<i>Anaxyrus americanus</i>	Bufonidae	20	1	4.8 (0–24.4)
<i>Hyla versicolor</i>	Hylidae	19	0	0 (0–19.8)
<i>Pseudacris crucifer</i>	Hylidae	2	0	0 (0–70.1)
<i>Pseudacris maculata</i>	Hylidae	1	1	50 (9.5–90.6)
<i>Lithobates pipiens</i>	Ranidae	14	7	33.3 (17.1–54.8)
<i>Lithobates septentrionalis</i>	Ranidae	13	3	18.8 (5.8–43.8)
<i>Lithobates sylvaticus</i>	Ranidae	78	22	22 (14.9–31.2)

tially, a phylogenetic component to *Bd* (Corey and Waite 2008) with some amphibian lineages showing greater susceptibility to chytridiomycosis than others.

The presence of *Bd* in multiple frog species in Itasca State Park highlights the pathogen's pervasiveness in North America. North American amphibian populations have been exposed to *Bd* since the early 1960s and have been found across the continent (Ouellet et al. 2005). Presence of *Bd* does not always lead to mortality or population declines (Retallick and Miera 2007) but, given its potential as a pathogen, the presence of the fungus is cause for further monitoring of infected populations.

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#### LITERATURE CITED

- ALTSCHUL, S. F., W. GISH, W. MILLER, E. W. MYERS, AND D. J. LIPMAN. 1990. Basic local alignment search tool. *J. Mol. Biol.* 215:403–410.
- ANNIS, S. L., F. P. DASTOOR, H. ZIEL, P. DASZAK, AND J. E. LONGCORE. 2004. A DNA-based assay identifies *Batrachochytrium dendrobatidis* in amphibians. *J. Wildl. Dis.* 40:420–428.
- BLAUSTEIN, A. R., AND J. M. KIESECKER. 2002. Complexity in conservation: lessons from the global decline of amphibian populations. *Ecol. Lett.* 5:597–608.
- COREY, S. J., AND T. A. WAITE. 2008. Phylogenetic autocorrelation of extinction threat in globally imperilled amphibians. *Divers. Distrib.* 14:614–629.
- HANKE, M., AND M. WINK. 1994. Direct DNA-Sequencing of Pcr-Amplified Vector Inserts Following Enzymatic Degradation of Primer and Dntps. *BioTechniques.* 17:858–860.
- HERO, J. M., S. E. WILLIAMS, AND W. E. MAGNUSON. 2005. Ecological traits of declining amphibians in upland areas of eastern Australia. *J. Zool.* 267:221–232.
- JOHNSON, M. L., AND R. SPEARE. 2003. Survival of *Batrachochytrium dendrobatidis* in water: Quarantine and disease control implications.

- Emerg. Infect. Dis. 9:922–925.
- KRIGER, K. M., AND J. M. HERO. 2007. The chytrid fungus *Batrachochytrium dendrobatidis* is non-randomly distributed across amphibian breeding habitats. *Divers. Distrib.* 13:781–788.
- LIPS, K. R., F. BREM, R. BRENES, J. D. REEVE, R. A. ALFORD, J. VOYLES, C. CAREY, L. LIVO, A. P. PESSIER, AND J. P. COLLINS. 2006. Emerging infectious disease and the loss of biodiversity in a Neotropical amphibian community. *Proc. Natl. Acad. Sci. U. S. A.* 103:3165–3170.
- , J. D. REEVE, AND L. R. WITTERS. 2003. Ecological traits predicting amphibian population declines in Central America. *Conserv. Biol.* 17:1078–1088.
- LONGCORE, J. R., J. E. LONGCORE, A. P. PESSIER, AND W. A. HALTEMAN. 2007. Chytridiomycosis widespread in anurans of northeastern United States. *J. Wildl. Manage.* 71:435–444.
- MUTHS, E., P. S. CORN, A. P. PESSIER, AND D. E. GREEN. 2003. Evidence for disease-related amphibian decline in Colorado. *Biol. Conserv.* 110:357–365.
- OUELLET, M., I. MIKAELEIAN, B. D. PAULI, J. RODRIGUE, AND D. M. GREEN. 2005. Historical evidence of widespread chytrid infection in North American amphibian populations. *Conserv. Biol.* 19:1431–1440.
- PEARL, C. A., E. L. BULL, D. E. GREEN, J. BOWERMAN, M. J. ADAMS, A. HYATT, AND W. H. WENTE. 2007. Occurrence of the amphibian pathogen *Batrachochytrium dendrobatidis* in the Pacific Northwest. *J. Herpetol.* 41:145–149.
- RETALICK, R. W. R., AND V. MIERA. 2007. Strain differences in the amphibian chytrid *Batrachochytrium dendrobatidis* and non-permanent, sub-lethal effects of infection. *Dis. Aquat. Org.* 75:201–207.
- SAS. 2007. JMP. SAS Institute Inc, Cary, North Carolina, USA.
- SIMMONS, J. E. 2002. *Herpetological Collecting and Collections Management*, Revised Edition. SSAR, Shoreview, MN.
- STEINER, S. L., AND R. M. LEHTINEN. 2008. Occurrence of the amphibian pathogen *Batrachochytrium dendrobatidis* in Blanchard's cricket frog (*Acris creptians blanchardi*) in the U.S. Midwest. *Herpetol. Rev.* 39:193–196.
- WEBB, R., L. BERGER, D. MENDEZ, AND R. SPEARE. 2005. MS-222 (tricaine methane sulfonate) does not kill the amphibian chytrid fungus *Batrachochytrium dendrobatidis*. *Dis. Aquat. Org.* 68:89–90.
- WOODHAMS, D. C., A. D. HYATT, D. G. BOYLE, AND L. A. ROLLINS-SMITH. 2008. The northern Leopard frog *Rana pipiens* is a widespread reservoir species harboring *Batrachochytrium dendrobatidis* in North America. *Herpetol. Rev.* 39:66–68.

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## Chytridiomycosis in Woodhouse's Toad (*Anaxyrus woodhousii*) in Colorado

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As scientists in the 1990s became aware that amphibian populations were experiencing population declines on a number of continents, the most perplexing mortalities were those observed in relatively pristine environments in which man-made habitat destruction, introduction of invasive species, or direct application of toxicants were not evident. Mass mortalities of amphibians in

these relatively untouched environments, such as wilderness areas of the American West and tropical rainforests in Australia and Central America, shared a number of similarities that suggested a common proximate cause. These similarities were: 1) mass mortalities were observed principally in metamorphosed amphibians; 2) populations experienced severe declines in size, and, in some cases, total extinction; 3) only some of the amphibian species in a given locality experienced declines; and 4) population declines were more pronounced at higher elevations or in cooler regions than in lower/warmer areas (Lips 1998; Carey et al. 1999). Many of the mass mortalities sharing this pattern have now been attributed to a chytrid fungal pathogen, *Batrachochytrium dendrobatidis* (hereafter *Bd*; Berger et al. 1998; Skerratt et al. 2007). The observations that *Bd* is more likely associated with mass mortalities of amphibians at higher elevations in mountain ranges than in the adjacent lowlands have been replicated on several continents. For instance, numerous mass mortalities have been noted above approximately 400 m in Costa Rica and Panama (Lips 1998; Lips 1999; Young et al. 2001) but, even though *Bd* has been documented histologically on a number of species living at lower elevations in these countries, no mass mortalities have been observed (Puschendorf et al. 2006). Die-offs are most pronounced at elevations over 400 m in Australia (McDonald and Alford 1999 and above about 2000 m in Spain (Bosch et al. 2001; Bosch and Martinez-Solano 2006).

Woodhouse's Toads (*Anaxyrus woodhousii*) (formerly *Bufo woodhousii*; Crother 2008) are widely distributed in the US throughout the western central plains and Rocky Mountain states, from Montana and South Dakota south to Arizona and Louisiana. In Colorado, this species generally occurs at lower elevations up to about 2100 m. It is replaced at higher elevations by Western Toads (*Anaxyrus boreas*) (formerly *Bufo boreas*; Crother 2008) which range from about 2500–3550 m (Livo and Yackley 1997). Only one area of sympatric occurrence of the two species has been reported in Colorado (Hammerson 1999; Harris 1963). Western Toads are very susceptible to *Bd* (Carey et al. 2006). Beginning in the 1970s, Western Toad populations have experienced numerous mass mortalities that have extirpated breeding populations from about 85–90% of their historical habitats (Carey 1993; Corn et al. 1989). Some of these mass mortalities have been attributed to *Bd* (Carey et al. 1999; Muths et al. 2003). In a few instances, Woodhouse's Toad carcasses have been observed in the wild, but the deaths could not be attributed to *Bd*. Some carcasses were observed during one study in Arizona, but the carcasses were too decomposed to analyze for the presence or absence of *Bd* (Sredl 2000). Additionally, one of us (LL) found a dead Woodhouse's Toad in Canyonlands National Park, Utah (17 August 1992, ca. latitude 38.06814, longitude 109.76871) and two dead Woodhouse's Toads at Colorado National Monument, Colorado (24 June 1999, latitude 39.01993, longitude 108.64672). Toads at both of these locations were very decomposed and observed before a PCR test for *Bd* was developed. However, to our knowledge, no observations of mass mortalities of Woodhouse's Toads attributed to *Bd* have been recorded in the literature (see Sullivan 2005). Therefore, the goal of this study was to determine whether or not *Bd* is present on Woodhouse's Toads in Colorado.

A total of 86 Woodhouse's Toads were collected at a number of sites, the elevations of which ranged from 1096 to 2032 m (Table 1). Elevation for each sampling site was determined by plot-